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Registry No. 3a, 83313-92-2; 3b, 83313-93-3; 5, 83313-94-4; (E)-6, 83313-95-5; dicyclopropyl ketone acetylhydrazone, 83313-96-6; dicyclopropyl ketone propionylhydrazone, 83313-97-7; 1-acetoxyethylidene, 83313-98-8; trans-1,2-diacetoxy-1,2-dimethylcyclopropane, 67079-75-8; 1-acetoxy-2-methoxy-1,2-dimethylcyclopropane, 83313-99-9; acetylhydrazine, 1068-57-1; dicyclopropyl ketone, 1121-37-5; acetyl chloride, 75-36-5; biacetyl, 431-03-8; acetic anhydride, 108-24-7; propionyl chloride, 79-03-8; 2,3-pentanedione, 600-14-6; 2-acetoxypropene, 108-22-5; 2-methoxypropene, 116-11-0; CH₃COOCCH₂CH₃, 83350-10-1.

Fluoranthene: Synthesis and Mutagenicity of Four Diol Epoxides

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The syntheses of diol epoxides 4a,b and 5a,b of the mutagenic hydrocarbon fluoranthene (1) are described. Standard methodology is applied to the synthesis of targets 4a,b but fails for the synthesis of 5a,b. The latter two diol epoxides can be assembled by a route utilizing stereoselective, directed epoxidations. Simple molecular orbital calculations have been used to predict the reactivity of the diol epoxides in their opening to triol carbocations. Diol epoxides 4a,b are predicted to be substantially more reactive than isomers 5a,b. The more reactive pair, 4a,b, may yield carbocations capable of alkylating cellular genetic material. This prediction is borne out in terms of the relative mutagenicity of the diol epoxides in a bacterial screen.

Fluoranthene (1, Chart I) occurs in various fossil fuels and combustion effluents at concentrations considerably greater than those of the most frequently studied polycyclic aromatic hydrocarbon, benzo[a] pyrene.² Further, we have found fluoranthene and benzo[a] pyrene to be approximately equipotent as mutagens for Salmonella typhimurium^{3a} and as mutagens for human lymphoblasts.^{3b} We have sought the syntheses of potential oxidative metabolites of fluoranthene to help identify the mode(s) of in vivo metabolism of the hydrocarbon and the structure(s) of the covalent adducts presumably formed from the activated hydrocarbon and DNA. Herein we outline our progress directed toward these ends. Specifically, we detail the syntheses and mutation assay of six possible metabolites of fluoranthene, the dihydrodiols 11 and 23 (see Schemes I and II), and the diol epoxides 4a,b and 5a,b (Chart I).

Strategy

Extensive study of the metabolic activation of polycyclic aromatic hydrocarbons, such as $benzo[a]pyrene,^{4a}$ has



syn-lsomers: epoxide oxygen syn to benzylic (allylic) OH, e.g., 4a and 5a anti-isomers: epoxide oxygen anti to benzylic (allylic) OH, e.g., 4b and 5b

identified the diol epoxides (see 2a,b, Chart I) as agents responsible for the alkylation of cellular macromolecules. The genesis of the diol epoxides occurs through oxidation of the parent hydrocarbon to an arene oxide, followed by hydration of the oxide to the corresponding trans-dihydrodiol, and oxidation of the dihydrodiol to one or both

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 Table I.
 Triol Carbocation Delocalization Energies^a

 and Carcinogenicities^b for Fluoranthene



^a $\Delta E \pi / \beta$ (HMO calculation) and $\Delta E_{deloc/\beta}$ (PMO calculation) are calculated values for the delocalization energy gained upon opening of a diol epoxide to the depicted triol carbocation. The HMO and PMO methods are π -system calculations only and methods do not take the hydroxyl substituents or the σ framework into account. ^b Qualitative carcinogenicities for the parent hydrocarbons from ref 5a. ^c The name of the parent aromatic hydrocarbon is given under each triol carbocation. ^d Qualitative carcinogenicity for fluoranthene, see ref 7. ^e PMO calculations cannot be made for nonalternant π systems.

of the stereoisomeric diol epoxides. The identification of anti-diol epoxide 2a as a principal active metabolite of benzo[a]pyrene led Jerina et al.^{4b} to comment on the exceptional stability of the triol carbocations formed from bay-region diol epoxides (e.g., 2a,b). These workers and others^{5a-d} have correlated theoretical parameters of the bay-region carbocations with the carcinogenic or mutagenic potency of the parent hydrocarbons. It is proposed that the reactivity of a diol epoxide toward genetic material correlates with the difference in delocalization energy between the diol epoxide and its derived triol carbocation. At least in theory, a readily formed carbocation, formed in situ from the corresponding diol epoxide, should show poor discrimination between strong (e.g., glutathione) and weak (e.g., DNA) cellular nucleophiles.⁶ Such a correlation must be approximate, at best, when predicting the carcinogenicity of the parent polycyclic aromatic hydrocarbon (see Tables I and III in supplementary material), as other complex factors will ultimately govern the in vivo formation and detoxification of any given diol epoxide.^{5e} The molecular orbital technique may fail also in predicting the reactivity of a diol epoxide in a biological assay (see e.g., ref 4a) owing among other factors to the requirement for absorption and intact delivery of the potential mutagen (carcinogen) to a crucial cellular target. Nonetheless, simple molecular orbital techniques may be of value in the initial evaluation of several potential metabolites of an aromatic hydrocarbon. Diol epoxides formed within a cell and capable of readily forming triol carbocations will likely



react rapidly with cellular macromolecules. These diol epoxides and their dihydrodiol precursors are good candidates for synthesis and biological evaluation.

Given the number of polycyclic aromatic hydrocarbons that appear to be activated as the diol epoxides,^{4a} we have sought the syntheses of the diol epoxides of fluoranthene. The symmetry of the hydrocarbon (1) limits the number of potential diol epoxide metabolites to a small group, viz., 3a,b, 4a,b, and 5a,b (Chart I). Of these, only 5a,b can be evaluated by the perturbational molecular orbital (PMO) method, which has been applied to benzo[a]pyrene and other polycyclic aromatic hydrocarbons.^{4b} Diol epoxides 3a,b and 4a,b appear to be structurally related to the bay-region, benzo[a] pyrene diol epoxides (2a,b), but the PMO method is not applicable to their nonalternant π systems. We have examined all of the fluoranthene diol epoxides by using simple Huckel molecular orbital (HMO) theory; 5a,b has been examined as well by the PMO method. Table I shows the results of these calculations. Values of $\Delta E \pi / \beta$ and $\Delta E_{deloc} / \beta$ for several other polycyclic aromatic hydrocarbons are given in Table III (see supplementary material).

In Table I the third column gives the value for $\Delta E \pi / \beta$, the difference in delocalization energies between diol epoxide and its corresponding triol carbocation, calculated by the simple Huckel molecular orbital (HMO) method. The fourth column gives the value of $\Delta E_{deloc}/\beta$, the difference in delocalization energy between the diol epoxides 5a,b and their corresponding triol carbocation, calculated by the PMO method.^{4b} The values for $\Delta E \pi / \beta$ are consistently larger than $\Delta E_{deloc}/\beta$ over a series of polycyclic aromatic hydrocarbons (Table III). Figure 1 (supplementary material) shows the close linear correlation between the two calculated values for several alternant π systems. The values of $\Delta E \pi / \beta$ for the nonalternant fluoranthene carbocations suggest that diol epoxides 3a.b (entry B) and 4a,b (entry C) will be at least as reactive as the bay-region diol epoxides from other polycyclic aromatic hydrocarbons (Figure 1). Diol epoxides 5a,b are expected to be the least reactive of the fluoranthene metabolites. The routes to racemic diol epoxides 4a,b and 5a,b via their corresponding dihydrodiols are presented below.

Diol Epoxide Syntheses

Diol Epoxides 4a,b. Diastereomers 4a,b are prepared by the standard diol epoxide synthesis described by Jerina

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et al.⁸ (Scheme I). Details of our syntheses appear in the *Experimental Section*.

Diol Epoxides 5a,b. The syntheses of diastereomers **5a,b** by the standard route to diol epoxides⁸ requires dihydrofluoranthene 14 (Scheme II) as starting material. Olefin 14 is prepared from tetrahydrofluoranthene¹² (13) by a modification¹³ of the known hydroxylation/dehydration sequence.¹⁴ Prevost oxidation¹⁰ of 14, however, fails to produce the corresponding *trans*-tetrahydrobenzoate (cf. 9, Scheme I). An alternate route to **5a,b** therefore was sought.

Cis hydroxylation of 14 affords tetrahydrodiol 15 (87%). The *cis*-diol 15 is converted into α -hydroxy ketone 18 via dehydration (15 \rightarrow 16, 97%), silylation (16 \rightarrow 17, 96%), and oxidation (17 \rightarrow 18, 61–79%).¹⁵ Protection of the tertiary hydroxyl group of 18 (92%) and stereoselective phenylselenylation provides keto selenide 20 (50–61%). The desired trans relationship of the hydroxyl groups in the target diol epoxides is achieved by stereoselective (8:1, trans/cis) reduction of ketone 20 (80–90%, mixture of 21 and its *cis*-diol isomer).^{16a,b} Selenoxide fragmentation from

(13) KOH/18-crown-6 in benzene has also been used in the oxidation of fluorene to fluorenone, see: Gokel, G. W.; Durst, H. D. Synthesis 1976, 168.

| Table II. | Mutagenicity | of Polycyclic | Aromatic | | |
|---------------------------------|--------------|---------------|----------|--|--|
| Hydrocarbons and Metabolites to | | | | | |
| | Salmonella t | vnhimurium | | | |

| Saimonetta typnimartam | | | | | |
|-------------------------|------------------|---|---------------------------|--|--|
| compd | PMS ^a | significant ^b induced mutation | concn, ^c µM | | |
| £1 | 0 | _ | 50 | | |
| Iluoranthene (1) | Α | + | 5 | | |
| 1 | 0 | | 100 | | |
| benzo[<i>a</i>]pyrene | Α | + | 4 | | |
| 111 | 0 | + | 140 | | |
| ainyaroaioi 11 | Α | + | 30 | | |
| | 0 | + | 2.5 - 3.1 | | |
| diol epoxide 4a | Α | + | 13-33 | | |
| dial openide th | 0 | + | 0.3-1.0 | | |
| diol epoxide 4b | Α | + | 10-18 | | |
| dilara dua di al 00 | 0 | | 300 | | |
| ainyaroaioi 23 | А | - | 300 | | |
| dial an autole Ma | 0 | + | 270 | | |
| dioi epoxíde ba | Α | - | 300 | | |
| dial an avida Fh | 0 | - | 300 | | |
| dior choxing ap | А | - | 300 | | |

^a O, no rat liver post-mitochondrial supernatant (PMS) added to assay; A, aroclor-induced PMS added to assay. ^b -, no significant induced mutation observed; +, significant induced mutation based on the 99% upper confidence limit of the historical negative controls. ^c Number listed is the lowest concentration range of significant induced mutation for positive responses, highest concentration tested for negative responses.

21 affords the key intermediate, protected *trans*-dihydrodiol 22 (63%).

The hydroxyl-group directing effect used to advantage in Scheme I fails when applied to the peracid oxidation of *trans*-dihydrodiol 23 in which the hydroxyl groups are axially disposed.¹¹ Nonetheless, the stereoselective conversion of protected dihydrodiol 22 into the *syn*- (5a) and *anti*- (5b) diol epoxides is possible by use of a modified strategy. Epoxidation of 22 by *t*-BuOOH/VO(acac)₂¹⁷ yields a single epoxide, 24 (82%). Upon desilylation (quantitative) 24 affords *anti*-diol epoxide 5b. Deprotection and acetylation of 22 (92%) provides acetate alcohol 25. Epoxidation of 25 by *t*-BuOOH/VO(acac)₂ yields a single epoxide, 26. Upon deacetylation (82–90%) 26 affords *syn*-diol epoxide 5a.

Biological Evaluation

The mutagenic potential of fluoranthene and its dihydrodiols and diol epoxides was assayed by determining bacterial (*Salmonella typhimurium* TM677) resistance to the purine analogue, 8-azaguanine (8-AG), in the presence of each compound as described by Skopek et al.¹⁸ Normally, 8-AG enters the cell and is metabolized and incorporated into nucleic acids where the toxic effects of the analogue are manifested. However, mutants unable to transport or phosphoribosylate 8-azaguanine grow in the presence of 8-AG. The concentration of a given mutagen required to increase the frequency of cells capable of growth in the presence of 8-AG provides a measure of the mutagenicity of the compound.

The biological evaluation of fluoranthene and the fluoranthene dihydrodiols and diol epoxides is summarized in

⁽⁷⁾ Preliminary results have indicated that fluoranthene is tumorigenic in an infant mouse lung adenoma bioassay; Busby, W. F. (M.I.T., Department of Nutrition and Food Science), personal communication.

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⁽¹⁰⁾ Newman, M. S.; Beal, P. J. J. Am. Chem. Soc. 1950, 72, 5163. Gunstone, F. D. Adv. Org. Chem. 1960, 1, 122.

⁽¹¹⁾ The Henbest effect observed during the epoxidation of allylic alcohols by peracids is more strongly manifested in pseudoequatorial alcohols. For a discussion of the lower stereoselectivity observed with pseudoaxial alcohols, see: Lee, H.; Harvey, R. G. *Tetrahedron Lett.* 1981, 22, 1657.

⁽¹²⁾ Braun, J.; Manz, G. Chem. Ber. 1931, 63, 2608.

⁽¹⁴⁾ Streitweiser, A.; Suzuki, S. Tetrahedron 1961, 16, 153.

⁽¹⁵⁾ The Corey-Kim oxidation (see: (a) Corey, É. J.; Kim, C. U. Tetrahedron Lett. 1974, 287; (b) Corey, E. J.; Kim, C. U. J. Am. Chem. Soc. 1972, 94, 7586) and the Swern oxidation (see: (c) Mancuso, A. J.; Huang, S.-L.; (d) Swern, D. J. Org. Chem. 1978, 43, 2480; (e) Mancuso, A. J.; Brownfain, D. S.; Swern, D. Ibid. 1979, 44, 4148) give poor yields of α -hydroxy ketone 18. The three-step sequence of Scheme II is preferable to either direct oxidation method.

^{(16) (}a) The mixture of diol isomers is separated after the selenoxide fragmentation (at stage 22); see Experimental Section. (b) For the reduction of acyclic keto selenides see: Leonard-Coppens, A. M.; Krief, A. *Tetrahedron Lett.* 1976, 3227.

⁽¹⁷⁾ Sharpless, K. B.; Verhoeven, T. R. Aldrichimica Acta 1979, 12, 63.

^{(18) (}a) Skopek, T. R.; Liber, H. L.; Krolewski, J. J.; Thilly, W. G. *Proc. Natl. Acad. Sci. U.S.A.* **1978**, *75*, 410. (b) Skopek, T. R.; Liber, H. L.; Kaden, D. A.; Thilly, W. G. Ibid. **1978**, *75*, 4465.

Table II. For fluoranthene and each potential metabolite the mutation assay was performed in the presence and in the absence of the post-mitochondrial supernatant (PMS) of aroclor-induced rat livers.¹⁸ The PMS contains the metabolizing systems responsible for the oxidative conversion of polycyclic aromatic hydrocarbons into their mutagenic forms. A compound capable of alkylating nucleic acids without activation may induce mutations in the absence of PMS. By contrast, a compound requiring further oxidative elaboration, e.g., the conversion of a polycyclic aromatic hydrocarbon into one of its diol epoxides, will show a mutagenic effect only upon activation by the metabolizing system.

Fluoranthene is mutagenic only in the presence of PMS (Table II). Its mutagenic potency is comparable to that of benzo[a]pyrene tested under the same conditions. Mutagenic dihydrodiol 11 is more potent in the presence of PMS, suggesting a role for a diol epoxide (e.g., 4a,b) in the mutagenic activation of the parent hydrocarbon. Both diol epoxides 4a and 4b are potent mutagens in the absence of the metabolizing systems (PMS). The lower activity in the presence of PMS may reflect destruction of the diol epoxides by the rat liver metabolizing systems. Dihydrodiol 23 does not display mutagenic activity up to $300 \,\mu$ M concentration in the presence or absence of PMS. Of the diol epoxides 5a,b, the syn-diol epoxide (5a) displays weak mutagenic activity; the anti-diol epoxide (5b) is inactive even at high concentration.

Discussion

The mutagenic activity of diol epoxides 4a,b and 5a,b correlates well with the prediction of their reactivity by molecular orbital techniques (Table I). Triol carbocations should form more readily from diol epoxides 4a,b ($\Delta E \pi / \beta$ = 1.160) than from the isomeric diol epoxides 5a,b ($\Delta E \pi / \beta$ = 0.718). The former pair of diol epoxides also is the more mutagenic by a significant margin. Thus, the triol carbocation(s) shown in entry C of Table I becomes a good candidate(s) for the molecular species responsible for the covalent modification of cellular macromolecules by fluoranthene. In principle, the low activity of diol epoxides 5a,b could be due to inefficient delivery of the compounds to the appropriate site of mutagenic action. Destruction of **5a.b** by solvolysis in the biological milieu is unlikely, however, given the predicted relative reactivities (4a,b)5a,b, Table I) and the potent mutagenicity of 4a,b (Table II).

Conclusive testing of the involvement of diol epoxides 4a, b during the activation of fluoranthene and in the alkylation of cellular genetic material must await further biological studies in our laboratories. Recently we have found¹⁹ dihydrodiol 11^{20} as the major component of the fluoranthene metabolite profile formed by activation of the hydrocarbon in an in vitro system. This finding and the results of the present paper point suggestively to either diol epoxide 4a or 4b as the causative agent(s) of fluoranthene-induced mutation. The final details of the molecular basis of mutagenesis for fluoranthene must be learned through (a) the determination of structure for the fluoranthene-DNA adduct(s)²¹ and (b) the demonstration that such adducts are, in fact, premutagenic lesions.

Experimental Section

General Procedures. ¹H NMR spectra were recorded on a Perkin-Elmer R-24B (60 MHz), Varian T-60 (60 MHz), JEOL HF-90 Q (90 MHz), Bruker WM-250 (250 MHz), or Bruker WM-270 (270 MHz) NMR spectrometer. ¹³C NMR spectra were recorded on a Bruker WM-250 spectrometer at 62.8 MHz. Chemical shifts are reported downfield from tetramethylsilane (Me₄Si) on the δ scale. Infrared spectra were determined on a Perkin-Elmer 283 B spectrophotometer. Mass spectra were measured at 70 eV on a Varian MAT 44 mass spectrometer. High-resolution mass measurements were performed on a Mattauch-Herzog (Du Pont Instruments) CEC 110 B mass spectrometer. Elemental analyses were performed by Robertson Laboratories, Florham Park, NJ. Melting points were determined in open capillaries on a Thomas-Hoover Unimelt apparatus and are uncorrected.

All reactions were performed under an inert atmosphere unless otherwise specified. The following solvents were distilled before use: tetrahydrofuran (THF, from sodium/benzophenone ketyl), CH_2Cl_2 (from P_2O_5), MeOH (from Mg turnings), benzene (from CaH_2), pyridine (from CaH_2), dioxane (from CaH_2), and diisopropylamine (from CaH_2). Anhydrous *p*-toluenesulfonic acid was prepared from the hydrate by azeotropic distillation with benzene. Silver benzoate was dried over P_2O_5 in vacuo. Flash chromatography was performed according to Still et al.²²

Diastereomeric Alcohols 7. Ketone 6⁹ (0.368 g, 1.67 mmol) in dry MeOH (10 mL) at 0 °C was stirred overnight with NaBH₄ (0.196 g, 5.18 mmol). After evaporation of the solvent, the residue was purified by extraction $(EtOAc/H_2O)$ and flash filtration (silica gel, 5:1 CHCl₃/EtOAc), affording the mixture of diastereomeric alcohols 7 (0.312 g, 84%). The mixture was routinely used in the subsequent dehydration, but the two alcohols were separated by flash chromatography (vide supra) for individual characterization. Data for faster-eluting isomer: mp 136-138 °C (recrystallized from EtOAc/hexanes); ¹H NMR (250 MHz, CDCl₃) δ 1.43 (m, 1 H), 2.24 (m, 1 H), 2.37 (m, 1 H), 2.45 (m, 2 H), 3.61 (dd, 1 H), 5.05 (t, 1 H), 7.29 (m, 4 H), 7.48 (m, 1 H), 7.59 (m, 1 H), 7.71 (m, 1 H); ¹³C NMR (62.8 MHz, CDCl₃) δ 147.7 (s), 145.5 (s), 141.5 (s), 139.4 (s), 137.0 (s), 128.3 (d), 127.2 (d), 126.9 (d), 124.8 (d), 124.2 (d), 120.7 (d), 119.1 (d), 65.8 (d), 45.3 (d), 33.9 (t), 20.9 (t); IR $(CHCl_3)$ 3590, 1450, 1430 cm⁻¹; mass spectrum, m/e 222 (M⁺). Anal. Calcd for C₁₆H₁₄O: C, 86.44; H, 6.35. Found: C, 86.30; H, 6.59. Data for slower-eluting isomer: mp 92-94 °C (recrystallized from EtOH/H2O); ¹H NMR (250 MH2, CDCl3) & 1.44 (m, 1 H), 2.12 (m, 2 H), 2.51 (m, 1 H), 2.62 (m, 1 H), 3.84 (dd, 1 H), 4.99 (t, 1 H), 7.29 (m, 4 H), 7.47 (m, 1 H), 7.59 (m, 1 H), 7.71 (m, 1 H); 13 C NMR (62.8 MHz, CDCl₃) δ 147.8 (s), 146.6 (s), 141.5 (s), 139.7 (s), 136.4 (s), 128.1 (d), 127.1 (d), 126.9 (d), 125.2 (d), 124.1 (d), 120.6 (d), 119.3 (d), 69.3 (d), 45.0 (d), 34.8 (t), 26.6 (t); IR (CHCl₃) 3590, 1448, 1430 cm⁻¹; mass spectrum, m/e 222 (M⁺). Anal. Calcd for C₁₆H₁₄O: C, 86.44; H, 6.35. Found: C, 86.16; H, 6.58.

Dihydrofluoranthene 8. The diastereomeric mixture of alcohols 7 (0.339 g, 1.52 mmol) and anhydrous *p*-toluenesulfonic acid (0.026 g, 0.152 mmol) were refluxed in dry benzene (20 mL). Water was removed azeotropically into a Dean–Stark trap. After 2 h the solution was concentrated and flash chromatographed (silica gel, benzene), affording pure 8 (0.310 g, quantitative) as a pale-yellow, crystalline solid: mp 71–72 °C; ¹H NMR (250 MHz, CDCl₃) δ 2.03 (tt, 1 H), 2.89 (ddd, 1 H), 3.93 (dd, 1 H), 6.13 (ddd, 1 H), 6.66 (dd, 1 H), 6.99 (d, 1 H), 7.24–7.92 (m, 6 H); UV λ_{max} 260 nm; exact mass calcd for C₁₆H₁₂ (M⁺) 204.09390, found 204.092 86.

Tetrahydrobenzoate 9. Dry silver benzoate (AgOCOPh, 0.374 g, 1.64 mmol) and iodine (0.207 g, 0.817 mmol) in dry benzene (20 mL) were stirred until all the I_2 was consumed (color change from red to yellow).¹⁰ To this suspension was added dihydro-fluoranthene 8 (0.167 g, 0.818 mmol) and the mixture was stirred in the dark overnight. Filtration through Celite, evaporation of solvent, and flash chromatography (silica gel, 4:1, CH₂Cl₂/hexanes) afforded pure 9 (0.365 g, quantitative) as a white crystalline solid:

⁽¹⁹⁾ Babson, J. R.; Wogan, G. N.; Wattley, R. V.; Russo-Rodriguez, S.;
Rastetter, W. H.; Andon, B.; Thilly, W. G., paper in preparation.
(20) Presumably only one enantiomer of dihydrodiol 11 is formed

⁽²⁰⁾ Presumably only one enantiomer of dihydrodiol 11 is formed metabolically; no assignment of absolute configuration has been made to the metabolic product.

⁽²¹⁾ A covalent adduct of fluoranthene and DNA has been identified in our laboratories following incubation of tritiated fluoranthene and the PMS from arachlor-induced rat livers with DNA; ref 19.

mp 152–153 °C; ¹H NMR (250 MHz, CDCl₃) δ 1.76 (ddd, 1 H), 3.21 (ddd, 1 H), 4.17 (dd, 1 H), 6.09 (ddd, 1 H), 6.69 (d, 1 H), 7.28–8.08 (m, 17 H); ¹³C NMR (62.8 MHz, CDCl₃) δ 166.6 (s), 166.1 (s), 146.3 (s), 141.5 (s), 139.6 (s), 133.3 (d), 131.6 (s), 130.0 (d), 129.9 (d), 129.8 (d), 128.5 (d), 127.6 (d), 127.3 (d), 126.5 (d), 124.3 (d), 120.9 (d), 120.3 (d), 76.2 (d), 74.4 (d), 42.5 (d), 32.6 (t); IR (CHCl₃) 1712, 1600, 1449, 1372 cm⁻¹; mass spectrum, *m/e* 446 (M⁺). Anal. Calcd for C₃₀H₂₂O₄: C, 80.69; H, 4.97. Found: C, 80.47; H, 5.20.

Dihydroben zoate 10. Tetrahydrobenzoate **9** (0.437 g, 0.980 mmol) and o-chloranil (0.301 g, 1.23 mmol) in dry dioxane (20 mL) were refluxed overnight. Evaporation of solvent and flash chromatography (silica gel, 5% EtOAc/hexanes) afforded **10** (0.209 g, 48%) as a pale-yellow, crystalline solid: mp 146–148 °C; ¹H NMR (250 MHz, CDCl₃) δ 6.41 (t, 1 H), 6.76 (d, 1 H), 6.80 (d, 1 H), 7.26–7.52 (m, 10 H), 7.61 (dd, 1 H), 7.70 (t, 2 H), 8.01 (m, 4 H); ¹³C NMR (62.8 MHz, CDCl₃) δ 165.8, 165.7, 141.9, 138.0, 137.1, 135.7, 135.0, 133.1, 129.8, 129.7, 129.5, 128.4, 128.3, 127.3, 125.8, 122.6, 120.9, 120.2, 117.2, 73.9, 71.5; IR (CHCl₃) 1712, 1448, 1264, 1101, 1090 cm⁻¹. Anal. Calcd for C₃₀H₂₀O₄: C, 81.07; H, 4.54. Found: C, 81.27; H, 4.71.

trans -Dihydrodiol 11. Dihydrobenzoate 10 (0.082 g, 0.185 mmol) and sodium methoxide (0.41 mL of 1 N solution in dry MeOH) were stirred at -20 °C in dry THF (5 mL). After 1 h, EtOAc (10 mL) and 1 N HCl (aq, 0.5 mL) were added. Extractive purification (EtOAc/H₂O) and flash chromatography (silica gel, 1:1 EtOAc/hexanes) gave 11 (0.036 g, 83%) as a pale-yellow solid: mp 167-168 °C dec; ¹H NMR (250 MHz, acetone- d_{θ}) δ 4.97 (dd, 1 H), 5.12 (d, 1 H), 6.61 (d, 1 H), 7.35 (m, 4 H), 7.59 (m, 1 H), 7.71 (m, 2 H); ¹³C NMR (62.8 MHz, acetone- d_{θ}) δ 143.0, 137.3, 137.0, 135.9, 135.6, 129.8, 129.6, 128.0, 126.6, 125.2, 123.2, 121.7, 119.6, 76.9, 75.4; IR (KBr) 3240, 1430, 1050, 1025, 950, 772, 752, 723 cm⁻¹; exact mass calcd for C₁₆H₁₂O₂: C, 81.34; H, 5.12. Found: C, 81.10; H, 5.16.

Bromo Triol 12. Dihydrodiol 11 (0.073 g, 0.310 mmol), *N*bromoacetamide (0.051 g, 0.372 mmol), and HCl (1 drop, aq 12 N) were stirred in 3:1 THF/H₂O (15 mL) for 2 h in the dark. Extractive purification (EtOAc/NaCl (aq, half satd), NaHCO₃ (aq, satd), NaCl (aq, satd)) and preparative TLC (silica gel, 3:7 hexanes/EtOAc) yielded 12 (0.101 g, 98%) as a pale-yellow solid: mp 97 °C dec; ¹H NMR (250 MHz, acetone- d_6) δ 2.05 (br s, 3 H), 4.63 (d, 1 H), 4.94 (dd, 1 H), 5.00 (d, 1 H), 7.24–7.43 (m, 4 H), 7.51–7.60 (m, 2 H), 7.66–7.70 (m, 1 H); IR (KBr) 3260, 1432, 1382, 1320, 1300, 1230, 1055, 1020, 1010, 990, 935, 920, 755, 730 cm⁻¹. Anal. Calcd for C₁₆H₁₃O₃Br: C, 57.68; H, 3.93. Found: C, 57.46; H, 3.91.

syn-Diol Epoxide 4a. Bromo triol 12 (0.054 g, 0.161 mmol) and freshly sublimed t-BuOK (0.018 g, 0.161 mmol) were suspended in dry Et₂O (5 mL) at 0 °C. After 30 min the mixture was filtered and the solvent removed in vacuo to afford 4a (0.040 g, quantitative) as a pale-yellow solid: mp 75–78 °C dec; ¹H NMR (250 MHz, acetone- d_6) δ 4.63 (m, 3 H), 7.33–7.74 (m, 5 H), 7.80 (d, 1 H), 7.90 (d, 1 H); IR (KBr) 3280, 1440, 1065, 1016, 1010, 913, 751, 742 cm⁻¹; exact mass calcd for C₁₆H₁₂O₃ (M⁺) 252.07864, found 252.07966.

anti-Diol Epoxide 4b. Dihydrodiol 11 (0.045 g, 0.191 mmol) and 99% *m*-chloroperoxybenzoic acid²³ (0.106 g, 0.611 mmol) in CHCl₃ (10 mL, passed through basic alumina) were stirred at 0 °C for 1 h. The mixture was purified by extraction (CHCl₃/ Na₂SO₃ (5% aq), NaHCO₃ (aq, half satd)) followed by liquid chromatography (Waters Radial-Pak, μ Bondapak CN Normal Phase, 3:7 EtOAc/hexanes), affording 4b (0.032 g, 66%) as a white solid: mp 126–129 °C dec; ¹H NMR (250 MHz, acetone-d₆) δ 3.88 (dd, 1 H), 4.39 (d, 1 H), 4.86 (d, 1 H), 7.32–7.52 (m, 5 H), 7.72 (m, 1 H), 7.87 (d, 1 H); ¹³C NMR (62.8 MHz, acetone-d₆) δ 143.6, 140.2, 138.3, 130.7, 130.6, 128.4, 124.2, 124.1, 122.1, 119.8, 76.5, 71.3, 61.2; IR (KBr) 3240, 1440, 1316, 1290, 1255, 1116, 1088, 1055, 1035, 890, 758, 730 cm⁻¹; mass spectrum, *m/e* 252 (M⁺). Anal. Calcd for C₁₆H₁₂O₃: C, 76.18; H, 4.79. Found: C, 76.38; H, 4.95.

Dihydrofluoranthene 14. A mixture of tetrahydrofluoranthene 13^{12} (6.91 g, 33.5 mmol), KOH (3.11 g, 55.5 mmol), and

1,4,7,10,13,16-hexaoxacyclooctadecane (18-crown-6, 1.14 g, 4.49 mmol) in benzene (45 mL) was stirred vigorously in an open Erlenmeyer flask for 24 h. Na_2SO_3 (60 mL aq, satd) and H_2O_3 (28 mL) were added to the orange suspension, and the mixture was stirred rapidly for 1 h. Extractive purification (benzene/ H_2O) and evaporation of solvent from the organic phase afforded an oil, which was dissolved in CH_2Cl_2 (200 mL) and treated with a catalytic amount of $MeSO_3H$. The reaction was monitored by TLC (silica gel, 4:1 hexanes/EtOAc) and additional MeSO₃H was added as needed to consume the starting material (a total volume of 5-8 drops of MeSO₃H used). Upon observing disappearance of starting material, the mixture was washed with NaCl (ag, half satd), dried $(MgSO_4)$, and concentrated in vacuo to provide the crude product. Flash chromatography (silica gel, 4:1 hexanes/ EtOAc) afforded crystalline 14 (5.50 g, 80%): mp 61-63 °C; ¹H NMR (60 MHz, CDCl₃) δ 2.4-3.3 (m, 4 H), 6.4 (t, 1 H), 6.8-7.7 (m, 7 H); mass spectrum, $m/e \ 204 \ (M^+)$.

cis-Diol 15. To a solution of dihydrofluoranthene 14 (5.49 g, 26.91 mmol) and N-methylmorpholine N-oxide dihydrate²⁴ (6.90 g, 45.1 mmol) in THF/H₂O (26:5, 31 mL) was added OsO₄ (0.143 g, 0.56 mmol) in THF (1 mL). The mixture was stirred for 6 days at ambient temperature and then NaHSO₃ (5.0 g) in H₂O (40 mL) was added, and the phases were vigorously stirred for ca. 15 min. Extractive purification (Et₂O/H₂O), filtration of the organic layer through Florisil, and evaporation of solvent provided the crude product as a solid. Trituration with CH₂Cl₂/Et₂O afforded pure 15 as a white crystalline solid (5.54 g, 87%): mp 171–173 °C; ¹H NMR (250 MHz, CDCl₃) δ 2.34 (s, 1 H), 2.36 (m, 2 H), 2.79 (m, 1 H), 3.03 (ddd, 1 H), 3.78 (dd, 1 H), 7.02–7.80 (m, 7 H); IR (CDCl₃) 3500–3100, 3030, 2950, 1605 cm⁻¹; mass spectrum, m/e 238 (M⁺). Anal. Calcd for C₁₆H₁₄O₂: C, 80.65; H, 5.92. Found: C, 80.53; H, 6.03.

Ketone 16. cis-Diol **15** (5.51 g, 23.17 mmol) was suspended in CH₂Cl₂ (100 mL) and MeSO₃H (3-5 drops) was added with stirring. When the mixture became homogeneous, the solvent was evaporated in vacuo, providing the crude, solid product. Recrystallization from CH₂Cl₂/hexanes and flash chromatography of the mother liquors (silica gel, 2:1 CH₂Cl₂/hexanes) afforded pure **16** as a white solid (4.94 g, 97%): mp 135–137 °C; ¹H NMR (250 MHz, CDCl₃) δ 2.34 (ddd, 1 H), 2.89 (ddd, 1 H), 3.06 (ddd, 1 H), 3.46 (m, 1 H), 4.45 (s, 1 H), 7.20–7.92 (m, 7 H); IR (CDCl₃) 3160, 2950, 1725, 1610, 1450 cm⁻¹. Anal. Calcd for C₁₆H₁₂O: C, 87.24; H, 5.49. Found: C, 86.96; H, 5.61.

α-Hydroxy Ketone 18. To a solution of ketone 16 (4.94 g, 22.45 mmol) and hexamethyldisilazane (5.7 mL, 27.0 mmol) in CH₂Cl₂ (90 mL) at -20 °C was added freshly distilled trimethylsilyl iodide (3.5 mL, 24.6 mmol). The mixture was allowed to warm to ambient temperature and stirred overnight. The mixture was diluted with CH₂Cl₂ (to a volume of ca. 200 mL) and centrifuged, and the supernatant was washed with ice-cold NaHCO₃ (aq, satd). The organic phase was dried (MgSO₄) and concentrated in vacuo to give pure, H₂O-sensitive silyl enol ether 17 (6.27 g, 96%): ¹H NMR (90 MHz, CDCl₃) δ 0.3 (s, 9 H), 2.7 (m, 2 H), 3.1 (t, 2 H), 7.0-7.9 (m, 7 H); mass spectrum, m/e 292 (M⁺).

A solution of 17 (4.79 g, 16.42 mmol) and 99% *m*-chloroperoxybenzoic acid²³ in CH₂Cl₂ (50 mL) was stirred at 0 °C for 35 min. Extractive purification (CH₂Cl₂/Na₂SO₃ (5% aq), NaHCO₃ (aq, satd)) and flash chromatography (silica gel, 10:1 CH₂Cl₂/ EtOAc) provided crystalline 18 (2.30 g, 61%): mp 152–154 °C; ¹H NMR (250 MHz, CDCl₃) δ 2.32 (s, 1 H), 2.45 (ddd, 1 H), 2.91 (ddd, 1 H), 3.10 (ddd, 1 H), 3.76 (ddd, 1 H), 7.14–7.80 (m, 7 H); IR (CDCl₃) 3560, 3500–3300, 3060, 2960, 2930, 1720 cm⁻¹; exact mass calcd for C₁₆H₁₂O₂ (M⁺) 236.083 73, found 236.082 20.

 α -Silyloxy Ketone 19. To a solution of α -hydroxy ketone 18 (1.06 g, 4.49 mmol) and hexamethyldisilazane (1.0 mL, 4.75 mmol) in pyridine (4.4 mL) was added trimethylsilyl chloride (0.70 mL, 5.50 mmol). The mixture was stirred at ambient temperature for 45 min. Extractive purification (CH₂Cl₂/NaHCO₃ (aq, satd)) and flash chromatography (silica gel, 1:1 CH₂Cl₂/hexanes) afforded crystalline 19 (1.26 g, 92%): mp 73–75 °C; ¹H NMR (250 MHz, CDCl₃) δ –0.33 (s, 9 H), 2.36 (ddd, 1 H), 2.87 (ddd, 1 H), 3.04 (ddd, 1 H), 3.63 (ddd, 1 H), 7.07–7.72 (m, 7 H); IR (CDCl₃) 3050, 2960, 1720 cm⁻¹; mass spectrum, m/e 308 (M⁺). Anal. Calcd for

⁽²³⁾ Fieser, L. F.; Fieser, M. "Reagents for Organic Synthesis"; Wiley: New York, 1967; Vol. 1, p 135.

C₁₉H₂₀O₂Si: C, 73.98; H, 6.54. Found: C, 73.83; H, 6.80.

Keto Selenide 20. To a cooled (-78 °C) solution of lithium diisopropylamide (10.4 mmol, prepared from 6.5 mL of 1.6 M n-BuLi and 1.5 mL of diisopropylamine) in THF (10.0 mL) was added over 10 min a solution of α -silvloxy ketone 19 (3.10 g, 9.90 mmol) in THF (6.0 mL). The resulting enolate was stirred for 30 min at -78 °C and quenched with trimethylsilyl chloride (2.50 mL, 19.82 mmol). The reaction was stirred an additional 30 min with continued cooling and then a solution of $PhSeCl^{25}$ (2.80 g, 14.66 mmol) in THF (4 mL) was added. The mixture was allowed to warm to ambient temperature and stirred for 5 h. Extractive purification (Et_2O/H_2O) and flash chromatography (silica gel, 2:1 CCl_4/CH_2Cl_2) gave pure 20 as an oily solid (2.20 g, 50%): ¹H NMR (250 MHz, CDCl₃) δ -0.32 (s, 9 H), 3.15 (dd, 1 H), 4.18 (dd, 1 H), 4.50 (dd, 1 H), 7.04-7.85 (m, 12 H); IR (CDCl₃) 3060, 2960, 2900, 1710, 1610 cm⁻¹; mass spectrum, m/e 464 (M⁺, ⁸⁰Se), 462 (M⁺, ⁷⁸Se).

Protected trans-Dihydrodiol 22. To a cooled (-15 °C) suspension of LiAlH₄ (0.244 g, 6.43 mmol) in Et₂O (30 mL) was added a solution of keto selenide 20 (1.95 g, 4.21 mmol) in Et₂O (18 mL, freshly distilled from LiAlH₄). The reaction was stirred for 10 min and quenched with a ground mixture of anhydrous Na₂SO₄ (6.98 g), H₂O (0.80 mL), and Celite (2.10 g). Dilution of the mixture with Et₂O (ca. 16 mL), filtration, and concentration of the filtrate afforded hydroxy selenide 21 as an oil (1.51 g, 80%): ¹H NMR (250 MHz, CD₂Cl₂) δ -0.458 (s, 9 H), 1.63 (s, 1 H), 2.75 (dd, 1 H), 3.48 (dd, 1 H), 4.47 (d, 1 H), 4.57 (dd, 1 H), 6.90-7.70 (m, 15 H).

A solution of hydroxy selenide 21 (1.51 g, 3.25 mmol) and 30% H_2O_2 (0.90 mL, 10.4 mmol) in THF (40 mL) at 0 °C was stirred for 1.5 h. The mixture was purified by extraction (Et₂O/H₂O, Na₂CO₃ (aq, satd)). Flash chromatography (silica gel, CH₂Cl₂) served to separate the minor *cis*-diol isomer (R_f 0.43) from the desired isomer 22 (R_f 0.29, crystalline solid, 0.612 g, 61%): mp 81-82 °C; ¹H NMR (250 MHz, CDCl₃) δ -0.45 (s, 9 H), 1.10 (s, 1 H), 4.48 (s, 1 H), 6.31 (dd, 1 H), 6.64 (d, 1 H), 6.99-7.60 (m, 7 H); IR (CDCl₃) 3580, 3420, 3040, 2960, 2900, 1640, 1600 cm⁻¹; exact mass calcd for C₁₉H₂₀O₂Si (M⁺) 308.123 30, found 308.1242.

Protected Diol Epoxide 24. To a solution of protected trans-dihydrodiol 22 (0.112 g, 0.360 mmol) and VO(acac)₂ (15.8 mg, 0.06 mmol) in dry benzene (1 mL) was added a solution of 6.5 M t-BuOOH (0.18 mL in CH₂Cl₂). After the mixture was stirred for 1 h at ambient temperature, Na₂SO₃ (5 mL, aq, half satd) was added. Extractive purification (benzene/H₂O) provided 24 (0.126 g, quantitative), which was routinely used in the subsequent step without purification (analytical TLC: trace base-line impurities). A small sample was purified by liquid chromatography (Waters Radial-Pak, µBondapak CN Normal Phase, hexanes) prior to characterization: ¹H NMR (270 MHz, CDCl₃) δ -0.50 (s, 9 H), 1.96 (d, 1 H), 4.01 (m, 2 H), 4.69 (dd, 1 H), 7.20-7.52 (m, 7 H); IR (CDCl₃) 3500, 3010, 2950, 1600 cm⁻¹; exact mass calcd for C₁₉H₂₀O₃Si (M⁺) 324.1182, found 324.1188.

anti-Diol Epoxide 5b. To a cooled (0 °C) solution of protected diol epoxide 24 (0.127 g, 0.390 mmol) in THF (3 mL) was added 1 M n-Bu₄N⁺F⁻ in THF (0.39 mL, 0.39 mmol). After being stirred for 10 min, the mixture was purified by extraction (Et₂O/H₂O) to give 5b as a pale-yellow solid (0.101 g, quantitative). For removal of residual colored impurities a small sample was rapidly flash chromatographed (silica gel, 80:20:1.5 CH₂Cl₂/EtOAc/NEt₃): mp 60 °C dec; ¹H NMR (270 MHz, CDCl₃) δ 2.14 (d, 1 H), 4.14 (m, 2 H), 4.85 (dd, 1 H), 7.32–7.62 (m, 7 H); IR (CDCl₃) 3560, 3510, 3060, 3020, 2960, 2930, 1600 cm⁻¹; exact mass calcd for C₁₆H₁₂O₃

(M⁺) 252.07864, found 252.0784.

trans-Dihydrodiol 23. A mixture of protected trans-dihydrodiol 22 (0.178 g, 0.580 mmol) and anhydrous K_2CO_3 (0.218 g, 1.58 mmol) in MeOH (0.70 mL) was rapidly stirred at ambient temperature for 3 h. Extractive purification (CH₂Cl₂/H₂O) afforded 23 as an off-white solid (0.138 g, quantitative) routinely used in the subsequent reaction without further purification. A small sample was purified by flash chromatography (silica gel, 4:1 CH₂Cl₂EtOAc) prior to characterization: mp 128 °C; ¹H NMR (270 MHz, CDCl₃) δ 1.21 (d, 1 H), 2.20 (s, 1 H), 4.66 (dd, 1 H), 6.38 (dd, 1 H), 6.77 (d, 1 H), 7.06–7.66 (m, 7 H); IR (CDCl₃) 3575, 3400, 3040, 2920, 1600 cm⁻¹; exact mass calcd for C₁₆H₁₂O₂ (M⁺) 236.083 76, found 236.0851.

Protected Diol Epoxide 26. To a solution of *trans*-dihydrodiol **23** (0.138 g, 0.59 mmol) in dry pyridine (0.60 mL) was added freshly distilled Ac₂O (0.06 mL, 0.63 mmol). The mixture was stirred at ambient temperature for 19 h and then purified by extraction (CH₂Cl₂/H₂O) and flash chromatography (silica gel, 6:1 CH₂Cl₂/EtOAc), affording pure acetoxy alcohol **25** (0.128 g, 78%) plus recovered starting material (19.8 mg). Data for **25**: R_f 0.63 (4:1 CH₂Cl₂/EtOAc); ¹H NMR (270 MHz, CDCl₃) δ 1.67 (s, 3 H), 2.03 (s, 1 H), 5.90 (d, 1 H), 6.25 (dd, 1 H), 6.84 (d, 1 H), 7.07-7.63 (m, 7 H).

To a solution of acetoxy alcohol **25** (0.118 g, 0.430 mmol) in dry benzene (1 mL) was added VO(acac)₂ (18.9 mg, 0.07 mmol) and 6.5 M t-BuOOH in CH₂Cl₂ (0.18 mL, 1.20 mmol). The reaction mixture was stirred at ambient temperature for 3 h and then additional 6.5 M t-BuOOH in CH₂Cl₂ (0.26 mL, 1.70 mmol) was added and stirring continued for 2 h. Extractive purification (benzene/Na₂SO₃ (aq, satd)) afforded **26** as a pale-yellow solid (0.127 g, quantitative): ¹H NMR (250 MHz, CDCl₃) δ 1.62 (s, 3 H), 3.74 (s, 1 H), 4.00 (dd, 1 H), 4.17 (d, 1 H), 6.25 (d, 1 H), 7.28–7.65 (m, 7 H); IR (CDCl₃) 3480, 3020, 2930, 2850, 1740 cm⁻¹; exact mass calcd for C₁₈H₁₄O₄ (M⁺) 294.0892, found 294.0918.

syn-Diol Epoxide 5a. To a cooled (-5 °C) solution of acetoxy alcohol 26 (0.075 g, 0.25 mmol) in MeOH (2.5 mL) was added anhydrous K_2CO_3 (3.9 mg, 0.28 mmol). The mixture was stirred for 10 min and then diluted with Et_2O and washed with H_2O . Evaporation of solvent from the organic phase and trituration with Et_2O afforded 5a as a white solid (50 mg, 80%): mp 148–150 °C; ¹H NMR (270 MHz, CDCl₃) δ 3.77 (s, 1 H), 4.02 (dd, 1 H), 4.14 (d, 1 H), 4.96 (s, 1 H), 7.34–7.67 (m, 7 H); IR (CDCl₃) 3570, 3470, 3060, 3020, 2970 cm⁻¹; exact mass calcd for $C_{16}H_{12}O_3$ (M⁺) 252.0786, found 252.0799.

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Registry No. 4a, 83349-66-0; **4b**, 83349-67-1; **5a**, 83291-40-1; **5b**, 83349-68-2; **6**, 38422-92-3; **7** (isomer 1), 83291-41-2; **7** (isomer 2), 83291-42-3; **8**, 35324-29-9; **9**, 83291-43-4; **10**, 83291-44-5; **11**, 82911-12-4; **12**, 83311-76-6; **13**, 20279-21-4; **14**, 30339-87-8; **15**, 83291-45-6; **16**, 83291-46-7; **17**, 83291-47-8; **18**, 83291-48-9; **19**, 83291-49-0; **20**, 83291-50-3; **21**, 83291-51-4; **22** (isomer 1), 83291-52-5; **22** (isomer 2), 83291-53-6; **23**, 83291-54-7; **24**, 83291-55-8; **25**, 83291-56-9; **26**, 83291-57-0.

Supplementary Material Available: Cation delocalization energies and carcinogenicities (Figure 1 and Table III (2 pages). Ordering information is given on any current masthead page.

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